Rabbit antibody to 200 Neurofilament (150-200)

| Code | OSN00108W |
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| ID Tag | Rb2588-230615-WS |
| Unit size | $100 \mu \mathrm{l}$ |
| Immunogen | A synthetic peptide from aa region 150-200 of human 200 Neurofilament conjugated to blue carrier protein was used as the antigen. The peptide is homologous in mouse and rat. |
| Conjugate | Unconjugated antibody |
| Also known | NF-H, Neurofilament triplet H protein, 200 kDa neurofilament protein |
| Host | NZ white rabbit |
| Purity | Whole serum |
| Clonality | Polyclonal |
| Isotype | Polyclonal, whole serum |
| Applications | IHC, WB. A dilution of $1: 1000$ to $1: 2000$ is recommended. The optimal dilution should be determined by the end user. Not yet tested in other applications. |
| Specificity | Specific for Neurofilament heavy polypeptide. |
| Spcs X-react. | Human, rat, mouse. Other species not yet tested. |
| Format | Lyophilised |
| Reconstitution | Reconstitute in $100 \mu \mathrm{l}$ of sterile water. Centrifuge to remove any insoluble material. |
| Storage | Maintain the lyophilised/reconstituted antibodies frozen at $-20^{\circ} \mathrm{C}$ for long term storage and refrigerated at $2-8^{\circ} \mathrm{C}$ for a shorter term. When reconstituting, glycerol (1:1) may be added for an additional stability. Avoid freeze and thaw cycles. |
| Expiry Date | 12 months after reconstitution |
| Shipping | This item will be shipped to you at ambient temperature in a lyophilised form. |
| Limitation | For research use only |



IHC-P on paraffin sections of mouse spinal cord.
The animal was perfused using Autoperfuser at a pressure of 110 mmHg with 300 ml $4 \%$ FA and further post fixed overnight before being processed for paraffin embedding. HIER: Tris-EDTA, pH 9 for 20 min using Thermo PT Module.
Blocking: $0.2 \%$ LFDM in TBST filtered thru $0.2 \mu \mathrm{~m}$.
Detection was done using Novolink HRP polymer from Leica following manufacturers instructions; DAB chromogen: Candela DAB chromogen from Osenses.
Primary antibody: dilution 1: 1000, incubated 30 min at RT using Autostainer.
Sections were counterstained with Harris Hematoxylin.

